

Reductive Enzyme Cascades for Valorization of Polyethylene Terephthalate Deconstruction Products

Madan R. Gopal, Roman M. Dickey, Neil D. Butler, Michael R. Talley, Daniel T. Nakamura, Ashlesha Mohapatra, Mary P. Watson, Wilfred Chen, and Aditya M. Kunjapur*



Cite This: *ACS Catal.* 2023, 13, 4778–4789



Read Online

ACCESS |

Metrics & More

Article Recommendations

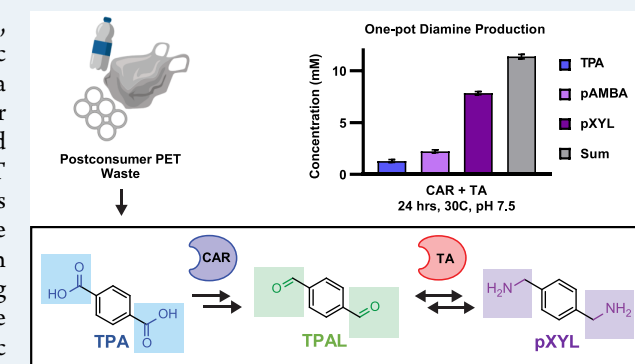
Supporting Information

ABSTRACT: To better incentivize the collection of plastic wastes, chemical transformations must be developed that add value to plastic deconstruction products. Polyethylene terephthalate (PET) is a common plastic whose deconstruction through chemical or biological means has received much attention. However, a limited number of alternative products have been formed from PET deconstruction, and only a small share could serve as building blocks for alternative materials or therapeutics. Here, we demonstrate the production of useful monoamine and diamine building blocks from known PET deconstruction products. We achieve this by designing one-pot biocatalytic transformations that are informed by the substrate specificity of an ω -transaminase and diverse carboxylic acid reductases (CAR) toward PET deconstruction products. We first establish that an ω -transaminase from *Chromobacterium violaceum* (cvTA) can efficiently catalyze amine transfer to potential PET-derived aldehydes to form monoamine *para*-(aminomethyl)benzoic acid (pAMBA) or diamine *para*-xylylenediamine (pXYL). We then identified CAR orthologs that could perform the bifunctional reduction of terephthalic acid (TPA) to terephthalaldehyde or the reduction of mono-(2-hydroxyethyl) terephthalic acid (MHET) to its corresponding aldehyde. After characterizing 17 CARs in vitro, we show that the CAR from *Segniliparus rotundus* (srCAR) had the highest observed activity on TPA. Given these elucidated substrate specificity results, we designed modular enzyme cascades based on coupling srCAR and cvTA in one pot with enzymatic cofactor regeneration. When we supply TPA, we achieve a $69 \pm 1\%$ yield of pXYL, which is useful as a building block for polymeric materials. When we instead supply MHET and subsequently perform base-catalyzed ester hydrolysis, we achieve $70 \pm 8\%$ yield of pAMBA, which is useful for therapeutic applications and as a pharmaceutical building block. This work expands the breadth of products derived from PET deconstruction and lays the groundwork for eventual valorization of waste PET to higher-value chemicals and materials.

KEYWORDS: plastics, polyethylene terephthalate, valorization, upcycling, reductive amination, carboxylic acid reductase, transaminase, biocatalysis

INTRODUCTION

The future of plastics production will require greater circularity to decrease dependence on petroleum as a raw material and to stem the ever-growing flow of plastic waste into landfills.^{1–5} Mechanical recycling of plastics has been the predominant technology to enable the reuse of postconsumer plastic waste. However, chemical deconstruction by synthetic or enzymatic approaches has emerged to allow for recovery of the monomer building blocks of various polymer plastics.^{6,7} While chemical deconstruction offers a simple path to remake the original, virginlike polymer, broader adoption and utility of chemical deconstruction will benefit from the design of sustainable methods to add value to deconstruction products, making them amenable to upcycled applications.^{3,8} One of the most widely used single-use plastics is polyethylene terephthalate (PET), a member of the thermoplastic category of polymers.⁹



Chemical and enzymatic deconstruction methods have been applied to PET to produce products such as bis(2-hydroxyethyl) terephthalic acid, mono(2-hydroxyethyl) terephthalic acid (MHET), and terephthalic acid (TPA).^{10–17} To date, most efforts have harnessed these compounds for either the resynthesis of PET or biological degradation as catabolized carbon sources or aliphatic building blocks derived from aromatic ring cleavage.^{18–20}

Received: December 16, 2022

Revised: February 13, 2023

Published: March 24, 2023



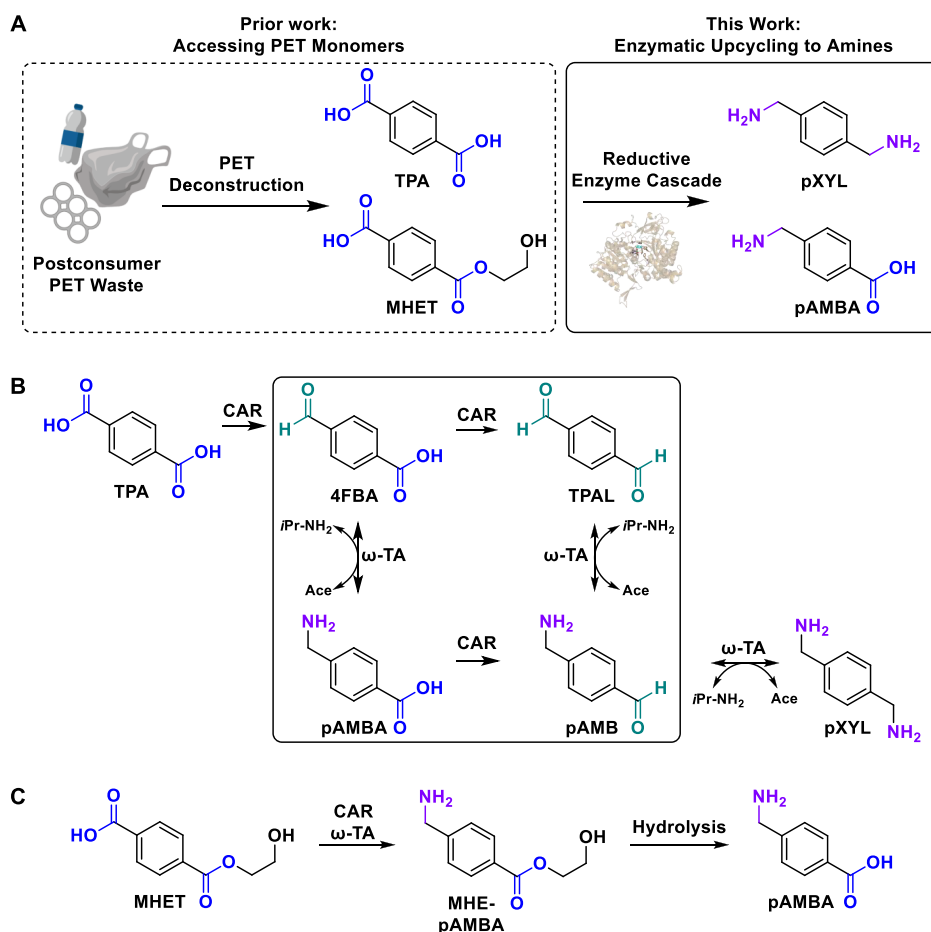


Figure 1. Strategies for valorizing deconstruction products of PET. (A) We envision using this enzyme cascade to upcycle PET waste by creating value-added amines for downstream application in therapeutics and polymer synthesis. (B) Envisioned reaction schematic for the one-pot reduction and amination of TPA to pXYL by an enzyme cascade consisting of CAR and ω -TA. The reaction cascade goes through four potential intermediate compounds, 4FBA, TPAL, pAMBA, and pAMB, due to successive reduction and amination reactions. (C) Substrate specificities of CAR and ω -TA could allow functionalization of MHET to pAMBA, a small-molecule therapeutic, by a simple one-pot enzymatic cascade. *iPr-NH₂*, isopropylamine; CAR, carboxylic acid reductase; ω -TA, ω -transaminase; Ace, acetone; TPA, terephthalic acid; 4FBA, 4-formylbenzoic acid; TPAL, terephthalaldehyde; pAMBA, *para*-(aminomethyl)benzoic acid; pAMB, *para*-aminomethylbenzaldehyde; pXYL, *para*-xylylenediamine.

We wondered whether we could design a route to convert PET deconstruction products to upcycled amines as alternatives to these products while preserving the aromatic nature of the terephthalate monomer (Figure 1A).²¹ Diamines such as *para*-xylylenediamine (pXYL) are value-added monomers for both thermoset and thermoplastic polymers. pXYL can be a component of polyamides, polyimides, or nonisocyanate polyurethanes (Figure S1).^{22,23} Such materials would substantially expand the breadth of products derived from PET deconstruction if an environmentally friendly option were available for valorization. However, chemical synthesis of pXYL requires multiple steps, elevated temperatures and pressures, and strong organic solvents.^{24,25} While diamines can be used as value-added monomers, a significant opportunity exists for the valorization of PET-derived monomers to monoamines such as *para*-(aminomethyl)benzoic acid (pAMBA), an antifibrinolytic drug used to promote blood clotting and treat fibrotic skin conditions.²⁶ Monofunctional molecules are often challenging to make from bifunctional substrates (e.g., TPA), resulting in poor atom economy and low selectivity.²⁷ However, enzymatic PET deconstruction offers a previously untapped potential to leverage substrates like MHET, a unique carboxylate with an

ester “protecting” group, which could allow for monofunctionalization of terephthalate.

Biocatalysis presents an industrially established and green alternative for the synthesis of amines and diamines from carbonyl-containing precursors.²⁸ Here, we envisioned an enzyme cascade that could convert TPA to pXYL in one pot (Figure 1B), and we wondered whether it could be adjusted such that it could instead generate pAMBA (Figure 1C). Our retrosynthetic design focused on steps of aldehyde consumption and aldehyde generation.^{29,30} Pyridoxal 5'-phosphate (PLP)-dependent ω -transaminases (ω -TA, EC 2.6.1.x) have been previously applied for the conversion of aldehydes to amines using simple cosubstrates like isopropylamine (*iPr-NH₂*), though to our knowledge, these enzymes have not been reported to have activity on bifunctional aromatic aldehydes.^{31–33} As PET deconstruction liberates carboxylates rather than aldehydes, we hypothesized that carboxylic acid reductases (CARs, E.C. 1.2.1.30) may be able to catalyze the reduction of carboxylates like TPA to their corresponding dialdehydes (which for TPA is terephthalaldehyde or TPAL).^{34–36} CARs are multidomain and polyspecific enzymes that perform the desirable 2e⁻ reduction of carboxylic acids to aldehydes at the cost of ATP and NADPH.³⁷ Given

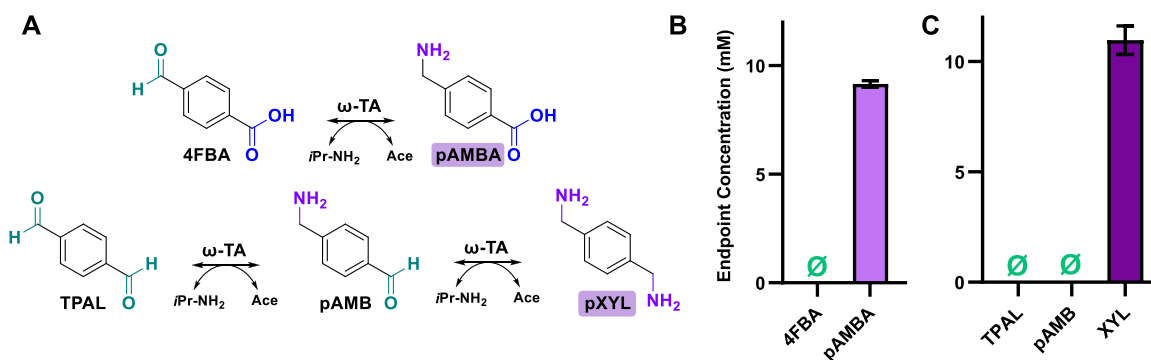


Figure 2. Testing cvTA on PET-derived aldehydes to produce amines. (A) cvTA reactions for the conversion of 4FBA and TPAL to their corresponding amines. Endpoint bioconversion of (B) 10 mM 4FBA to pAMBA and (C) 10 mM TPAL to pXYL by 1.5 μ M cvTA in 100 mM HEPES pH 7.5, 400 μ M PLP with 40 mM *iPr-NH₂* as the amine donor. This reaction proceeds to completion after 24 h at 30 $^{\circ}$ C, with no detection of aldehyde reactants or intermediates after 24 h. Sample size is $N = 3$, and all data are shown as the mean \pm standard deviation.

the broad substrate scope of both CAR and ω -TA families, some precedent for this type of cascade exists for certain aliphatic or heterocyclic chemistries.^{38,39} In 2019, Fedorchuk et al. designed a CAR and ω -TA cascade for one-pot transformation of the dicarboxylate adipic acid to hexamethylenediamine, a diamine precursor to nylon.⁴⁰ That study achieved only 30% yield, even after significant engineering to develop two CAR variants for use alongside two distinct transaminase orthologs. A major challenge was that each unique CAR or TA variant could accept a monofunctionalized intermediate or a bifunctionalized intermediate but not both.

Here, we report the design of enzyme cascades that feature single ω -TA and CAR variants to produce amines at high selectivity and yield in a one-pot reaction from PET-derived monomers. We examined the specificity of the ω -TA from *Chromobacterium violaceum* (cvTA), which successfully accepted several PET-derived aldehydes while harnessing *iPr-NH₂* as an amine donor. We next explored the diversity of natural CARs using a protein sequence similarity network (SSN) to identify known and novel putative CARs with activity on TPA. Our study revealed unexpected substrate specificity that enabled us to design highly selective routes to pXYL or to pAMBA by coupling the CAR from *Segniliparus rotundus* (srCAR) to cvTA. Our work represents a notable advance in the green conversion of PET deconstruction products to aromatic (di)aldehydes and (di)amines that could serve as platform intermediates for value-added polymeric materials or pharmaceuticals. By doing so, our work expands the sparse list of biological tools currently available for valorization of plastic deconstruction streams.

RESULTS AND DISCUSSION

An ω -TA from *Chromobacterium violaceum* Accepts PET-Derived Aldehydes. From our retrobiosynthetic analysis, we hypothesized that ω -TAs could catalyze the synthesis of diamines and monoamines from PET-derived aromatic aldehydes. We chose cvTA because of its wide substrate scope, well-characterized reaction mechanism, retention of activity in organic cosolvents, and compatibility with the simple amine donor, isopropylamine (*iPr-NH₂*).^{33,41,42} After identifying the reactions in which the ω -TA could participate (Figure 2A), we expressed, purified, and tested cvTA activity on 4-formylbenzoic acid (4FBA) and TPAL using *iPr-NH₂* as a cosubstrate.

After addition of 10 mM 4FBA to the reaction buffer containing 1.5 μ M cvTA, 40 mM *iPr-NH₂*, 5% DMSO, 100 mM HEPES pH 7.5, and 400 μ M PLP, we observed complete conversion by HPLC-UV analysis after incubation for 24 h at 30 $^{\circ}$ C (Figure 2B). Under the same reaction conditions, we also supplemented 10 mM TPAL, which initially resulted in turbidity. However, we were pleased to see the reaction turn translucent at 24 h, and HPLC-UV analysis indicated complete conversion of 10 mM TPAL to the desired diamine pXYL (Figure 2C). The highly efficient conversion of candidate aldehyde substrates to amine products catalyzed by cvTA led us to next wonder if CARs could generate these aldehydes from PET deconstruction products.

Bioprospecting Known and Putative CARs for Activity on Potential PET Deconstruction Products.

While CARs are extremely useful enzymes, bioprospecting for CAR remains relatively underexplored, likely due to many factors including only recent elucidation of its structure and catalytic cycle, cost of synthesis and assembly of the >3500 bp CAR gene, and the need for stoichiometric equivalents of NADPH and ATP per catalytic cycle.^{37,43,44} To address this gap in the literature and to identify CARs with specificity and activity toward products of PET deconstruction, we sought to investigate the natural CAR landscape. We first created a protein SSN using the Enzyme Function Initiative Enzyme Similarity Tool⁴⁵ and populated the SSN by selecting five previously characterized CARs, with at least one from each of four phylogenetic types of CARs:⁴⁶ (i) type I CAR from *Nocardia iowensis* (niCAR);⁴⁷ (ii) type II CAR from *Stachybotrys bisbyi* (sbCAR);⁴⁸ (iii) type III CARs from *Neurospora crassa* (ncCAR) and *Thermothelomyces thermophilus* (ttCAR);^{49,50} (iv) type IV CAR from *Trametes versicolor* (tvCAR2).⁵¹

For each of these CARs, we performed standard protein BLAST against the NCBI nonredundant protein sequence database. We ultimately populated the SSN with 500 sequences and visualized it using Cytoscape,⁵² where we observed a clear distinction between bacterial CARs and fungal type II, III, and IV CARs. The SSN also shows clusters of related enzyme classes known as L-amino adipate semialdehyde dehydrogenases (E.C. 1.2.1.31)⁵³ and L-amino adipate reductases (E.C. 1.2.1.95) (Figure 3A).⁵⁴ Based on this representative network, we selected 22 putative CARs for cloning and

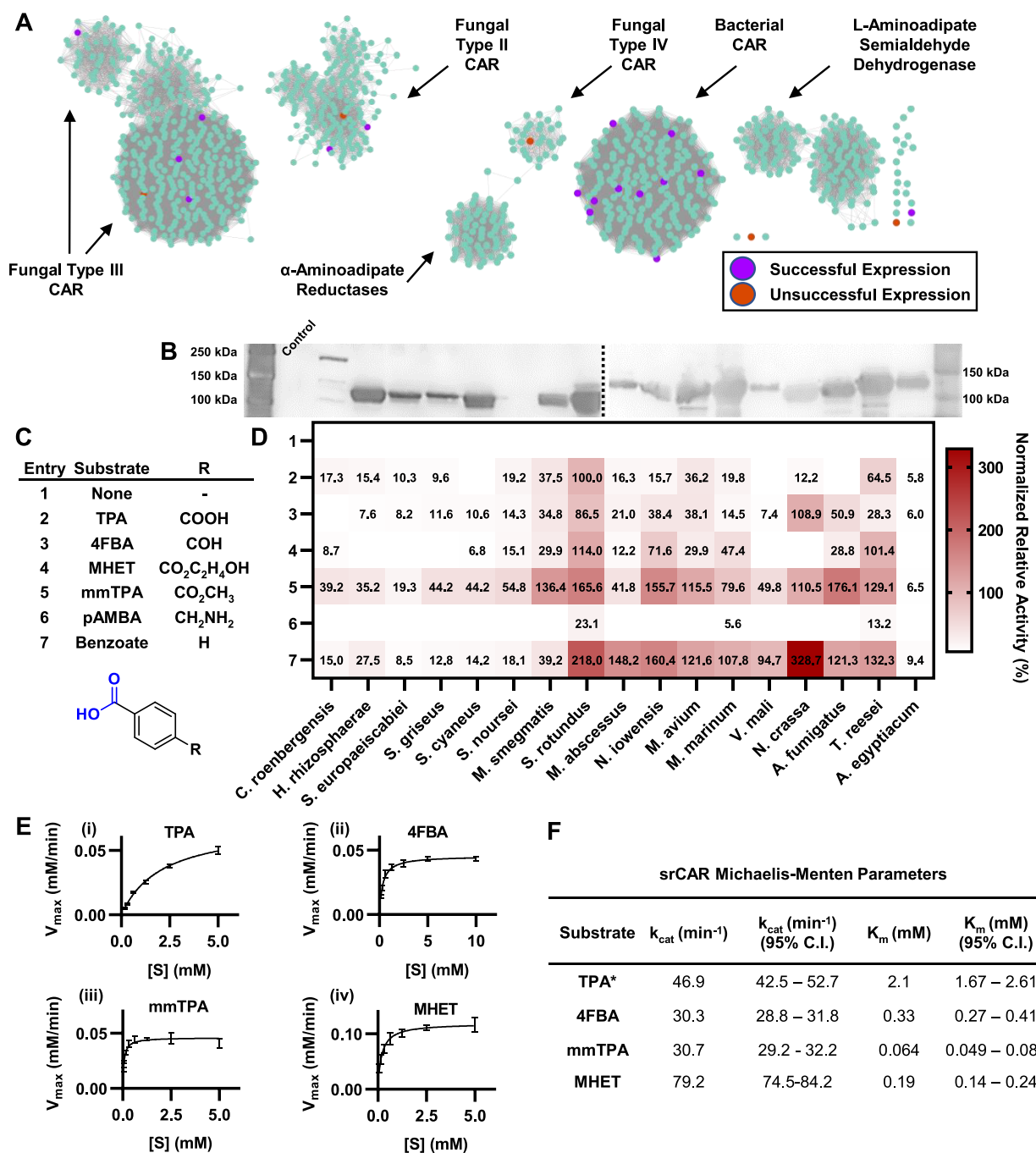


Figure 3. Evaluating CAR specificity on products of PET deconstruction. (A) Using an SSN, we sampled CARs across distinct protein clusters to probe for activity on TPA. Of 22 cloned CARs, 17 expressed, with 9 of those CARs being unreported in literature. (B) Anti-His₆ Western blot of cell lysate from *E. coli* BL21 (DE3) harboring different CARs is shown by phylogeny. CARs were expressed under optimized conditions. The Western blot is comprised of two separate blots, which are delineated by the dashed lines and contain their own protein ladders. The first lane of the Western blot is a control lane consisting of lysate from *E. coli* BL21 (DE3) with no CAR plasmid. A band corresponding to the putative CAR from *Streptomyces noursei* was not detected during the Western blot analysis but was confirmed in a separate experiment. (C) We evaluated the substrate specificity of CAR on TPA, 4FBA, MHET, and mmTPA, which are carboxylates related to PET depolymerization. In addition, we tested pAMBA as it is a potential reaction intermediate in our proposed multienzyme cascade for the formation of amines. (D) Under mild aqueous conditions, we tracked the conversion of CAR on TPA, 4FBA, MHET, mmTPA, pAMBA, and the model aromatic acid, benzoic acid. Here, the oxidation of NADPH is measured as a proxy to aldehyde formation as depletion of NADPH is linked to the consumption of the substrate. The heatmap was generated by calculating the triplicate average of initial rates of each CAR-substrate pair. The heatmap values represent the average initial rate normalized to the average activity of srCAR on TPA of $4.80 \text{ mM} \cdot \text{NADPH}_{ox} \cdot \text{h}^{-1} \cdot \mu\text{M}_{CAR}^{-1}$. CARs-substrate pairs with <5% normalized relative activity are not shown (E) Michaelis–Menten study was performed on the highest performing CAR from *Segniliparus rotundus* to evaluate k_{cat} and K_m on TPA (i), 4FBA (ii), MHET (iii), and mmTPA (iv). (F) Michaelis–Menten parameters and 95% confidence intervals were fitted using nonlinear least-squares regression on Graphpad Prism v9.3.1. Sample size is $N = 3$ for all numerical data. *srCAR kinetic parameters on TPA are reported as apparent values due to difficulty decoupling monofunctional and bifunctional reductions.

further investigation: 10 from the cluster occupied by the promiscuous type I bacterial CARs (5 previously uncharacterized); 3 from the presumed type II cluster (2 previously uncharacterized); 4 from the presumed type III cluster (2 previously uncharacterized); 1 putative enzyme from *Aspergillus fumigatus* from the less closely related portion of the type III CAR cluster; 1 putative enzyme from the presumed type IV cluster; and 3 nonclustered, uncharacterized putative enzymes from distinct eukaryotes (Table S1).

With the aid of the Joint Genome Institute, we cloned the 22 CARs chosen from the SSN into a vector containing the *sfp* gene from *Bacillus subtilis* (pZE/His-CAR-RBS2-*sfp*, see the Supporting Information Methods). Concomitant expression of CAR with *sfp*, which encodes a 4'-phosphopantetheinyl transferase (PPTase), allows for activation of the apo-CAR through the posttranslational addition of a phosphopantetheine arm.⁵⁵ This modification allows for the transfer of the bound thioester intermediate from the adenylation domain to the reduction domain during the catalytic cycle. When expressed under standard conditions, we noticed that several CARs exhibited poor expression. We optimized the media and expression temperature (Table S1) and were able to visualize the soluble expression of 17 CAR orthologs by performing SDS-PAGE (Figure S2) and a Western blot on the cell lysates (Figure 3B). We were then able to purify all 17 CARs using Ni-affinity chromatography.

After purifying our CAR enzymes, we investigated the in vitro activity of our purified CARs on potential PET deconstruction products, starting with TPA (Figure 3C). By adding 5 mM carboxylate stock in pure DMSO to a buffer containing a final concentration of 10 mM MgCl₂, 0.5 mM NADPH, 1 mM ATP, 100 mM HEPES pH 7.5, and 1.5 μM CAR (~0.2 mg/mL), we measured the initial rates of each CAR at 25 °C spectrophotometrically at 340 nm by tracking the oxidation of the reductive cofactor NADPH.⁵⁶ We found that our highest performing CAR on TPA was srCAR (from *S. rotundus*), which had an initial rate of $4.80 \pm 0.36 \text{ mM} \cdot \text{NADPH}_{\text{ox}} \cdot \text{h}^{-1} \cdot \mu\text{M}_{\text{CAR}}^{-1}$ (Figures 3D, S3). By normalizing the initial rates of all CAR orthologs using the activity of srCAR on TPA, we saw that trCAR (from *Trichoderma reesei*), msCAR (from *Mycobacterium smegmatis*), and mavCAR (from *Mycobacterium avium*) show 64.5, 37.5, and 36.2% normalized relative activity, respectively. During the preparation of this manuscript, the first example of a CAR that exhibits activity on TPA was reported.⁵⁶ This report demonstrated activity of mmCAR (from *Mycobacterium marinum*) on TPA in resting *Escherichia coli* whole cells, with no other CARs tested. Intrigued by that result, we performed our own analysis on mmCAR, albeit in an in vitro setting. We saw that mmCAR only exhibited 19.8% normalized activity on TPA relative to srCAR (Figure 3D). Curiously, mmCAR had very low activity in vitro on 4FBA, even at increasing concentrations of mmCAR (up to 4.5 μM), indicating that 4FBA may be a poor substrate (Figure S4). Analysis of CAR activity allowed us to characterize several “generalist” CARs, like srCAR, mavCAR, and msCAR, which had activity on both TPA and 4FBA. We also noticed unique “specialist” CARs in our heatmap, namely, from the fungal kingdom. Two of the top three CARs based on the activity on 4FBA were ncCAR (from *N. crassa*) and afCAR (from *A. fumigatus*), and yet they had minimal levels of activity on TPA. We were also pleasantly surprised by the breadth of reduction performed by trCAR, a previously untested putative aryl acid reductase, which showed

soluble expression and reduction activity on the full substrate scope.⁵⁷

We sought to further aid our cascade design by investigating whether CARs could accept an even broader range of starting molecules that could be derived from PET (Figure 3C). Based on the literature, we hypothesized that CARs may prefer aryl substrates that contain ring substituents with greater electron-withdrawing propensity.⁵⁸ Therefore, one compound of interest to us was monomethyl terephthalate (mmTPA). Encouragingly, here we succeeded in converting TPA to mmTPA at a yield of 76% using chemical synthesis (Supporting Information Methods). Additionally, we were able to convert dimethyl terephthalate, which is a product of methanolysis of PET, selectively to mmTPA at 89% yield (Supporting Information Methods).⁵⁸ A recent paper also shows that mmTPA can be produced from TPA with high selectivity using a carboxyl methyltransferase from *A. fumigatus*.⁵⁹ When we tested CAR activity on mmTPA, the general trend across the enzyme family was that CARs exhibited high activity on mmTPA. While srCAR only exhibited a 1.7-fold increase in initial activity on mmTPA when compared to its activity on TPA, niCAR (from *N. iowensis*) exhibited a noteworthy 9.9-fold increase in activity compared to its activity on TPA.

Another potential monoester/monoacid product of enzymatic deconstruction of PET is MHET. In fact, MHET is in some ways a more accessible enzymatic deconstruction product of PET, given that a MHETase is often the second step in published PETase/MHETase cascades that obtain TPA via MHET.⁶⁰ MHET is a particularly interesting candidate substrate given its large size relative to known substrates of CARs. Excitingly, niCAR, mmCAR, srCAR, and trCAR showed between 1.1 and 4.6-fold higher activity on MHET as compared to their respective activities on TPA. Overall, the results observed for the monoesters mmTPA and MHET were encouraging not just for the faster kinetics observed but also because they offered the potential to produce monoamine products with the esters serving akin to protecting groups.

When screening CARs against potential PET deconstruction products, we were also curious about activity on potential intermediates from our envisioned enzyme cascade for diamine production. The conversion of TPA to pXYL by a CAR and ω-TA cascade would theoretically result in a reaction fork, where 4FBA could either be reduced by CAR to TPAL or aminated by ω-TA to pAMBA, an amine-containing aromatic carboxylate (Figure 1B). When we provided pAMBA as a candidate substrate, srCAR and trCAR showed 23.1 and 13.2% normalized relative activity, respectively, with mmCAR showing detectable but very poor activity of 5.6% normalized relative activity. The low activity of CARs on pAMBA presented potential drawbacks and benefits—it could be a bottleneck on route to pXYL, but it could increase the likelihood of selectively producing pAMBA when that is the desired product.

To better understand the kinetics of our highest performing CAR, we sought to characterize k_{cat} and K_{m} of srCAR on TPA, 4FBA, mmTPA, and MHET as shown by the Michaelis–Menten plots (Figure 3E). Using the previously described plate reader-based NADPH oxidation assay, we observed that the average $k_{\text{cat,apparent}}$ and $K_{\text{m,apparent}}$ on TPA is 46.9 min^{-1} (95% C.I. 42.5–52.7 min^{-1}) and 2.1 mM (95% C.I. 1.7–2.7 mM), respectively (Figure 3F). We report these values as “apparent” as srCAR is a promiscuous enzyme and catalyzes not only the

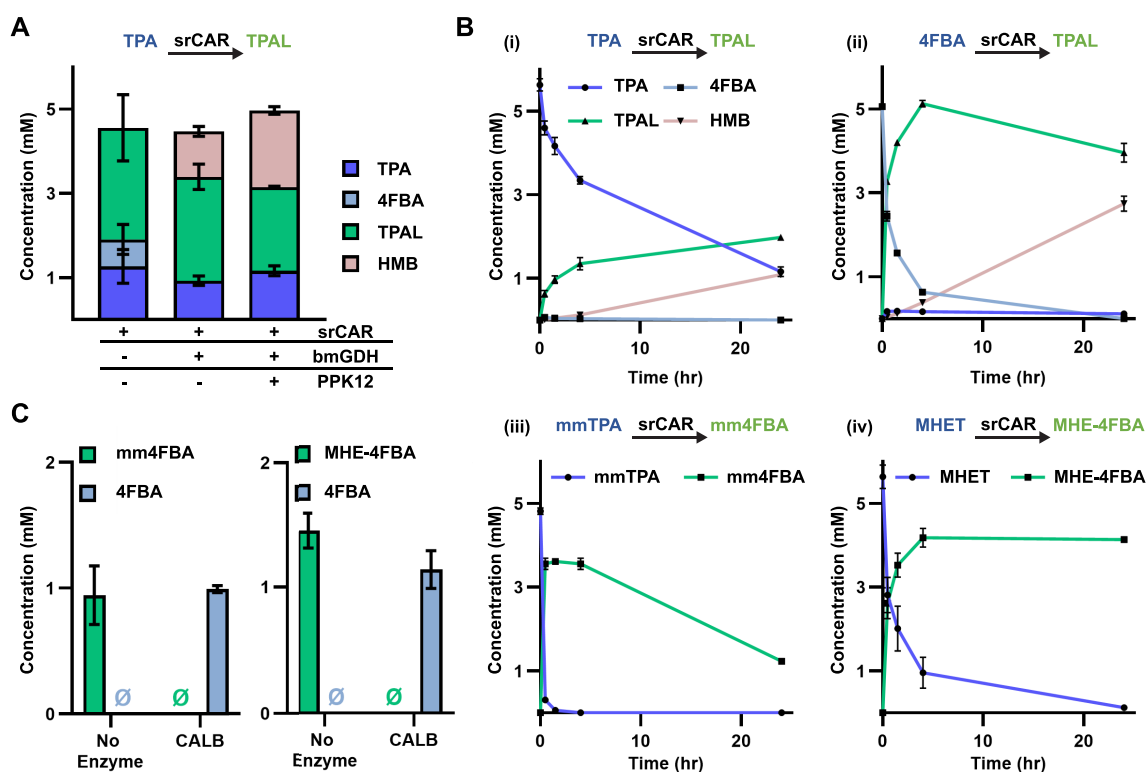


Figure 4. Reduction of PET-derived deconstruction products catalyzed by srCAR. (A) Endpoint in vitro assays showing the conversion of 5 mM TPA to TPAL after 24 h with and without cofactor regenerating enzymes. Our multienzyme cascade shows the overreduction of TPA to an alcohol, HMB. (B) Time course analysis of the enzymatic reduction of (i) 5 mM TPA, (ii) 5 mM 4FBA, (iii) 5 mM mmTPA, and (iv) 5 mM MHET by CAR. All experiments included a three-enzyme NADPH and ATP regeneration system. Reduction of TPA and 4FBA shows overreduction of aldehydes to alcohols. While mmTPA shows a decline in concentration over time to the purported alcohol product, we were not able to independently verify this. To our surprise, MHET shows no overreduction to the alcohol. (C) At our chosen endpoint of 24 h, lipase from *Candida antarctica* (CALB) completely de-esterified 1 mM of both MHE-4FBA and mm4FBA at 30 °C, providing an alternate route to forming aldehydes from PET-derived esters. Sample size is $N = 3$, and all data are shown as the mean \pm standard deviation. Error bars for certain points of the bioconversion of mmTPA to mm4FBA are smaller than the height of the symbol.

single reduction of TPA to 4FBA but also the subsequent reduction of 4FBA to TPAL, which makes it challenging to quantify biochemical parameters for only the first reaction. Fortunately, we can analyze just the second reaction in isolation by supplying 4FBA. We observed that the average k_{cat} and K_m of srCAR on 4FBA is 30.3 min^{-1} (95% C.I. 28.8 to 31.8 min^{-1}) and 0.33 mM (95% C.I. 0.27–0.41 mM), respectively. We also determined the srCAR kinetic parameters for MHET and mmTPA, which interestingly exhibit approximately 1 or 2 orders of magnitude lower K_m , respectively, than TPA, allowing for higher catalytic efficiency on these substrates (Figure 3F).

Sequence and Structural Insights of Natural CARs through Homology Modeling and Molecular Docking.

To explore whether the properties of active sites of predicted protein structures might correlate with the experimentally observed activity, we performed computational analyses. We first generated predictions of the 3D structure of the CAR adenylation domains (A-domains) using AlphaFold 2.1.⁶¹ Predicted structures are deposited as PDB files in Github (<https://github.com/KunjapurLab/CAR-Predicted-Structures.git>). For niCAR, the previously generated crystal structure was used instead of a prediction (PDB code: 5MSC).⁴³ AlphaFold did not generate a high confidence structure for the putative CAR from *Cafeteria roenbergensis*; thus, we omitted this protein from further analysis. We used the predicted structures to

compare binding pocket residues, binding pocket size, and substrate interactions via molecular docking.⁶² We identified binding pocket residues based on previous substrate-bound crystal structures and highly conserved signature sequences in the catalytic domains by multiple sequence alignment using Clustal Omega and Jalview.^{46,63} From this analysis, we found that the binding pocket residues are either highly conserved or vary with no clear pattern unique to our best experimentally observed enzymes (Figure S5). However, when we analyzed the average bottleneck radius using the CAVER webtool, we see that several of the CARs that are highly active on our substrate panel have larger predicted adenylation domain binding pocket radii (Figure S6A).⁶⁴

We then explored molecular docking on PET deconstruction products using the AlphaFold predicted protein structures and Autodock Vina. Docking analysis predicted that TPA could bind the predicted enzyme binding pocket of srCAR (Figure S6B). However, molecular docking did not show any observable trend matching that of our experimentally obtained result. Prior studies have suggested that the ϵ -nitrogen atom of a conserved lysine binding pocket residue can form hydrogen bonds with the acid acyl-AMP complex, particularly with the ribose-ring oxygen atom, and that the distance (d1) between the nitrogen atom and the oxygen atom of the Lys residue and ribose ring, respectively, could be a nimble indicator of CAR activity (Figure S6C).^{65,66} However, we did not see a

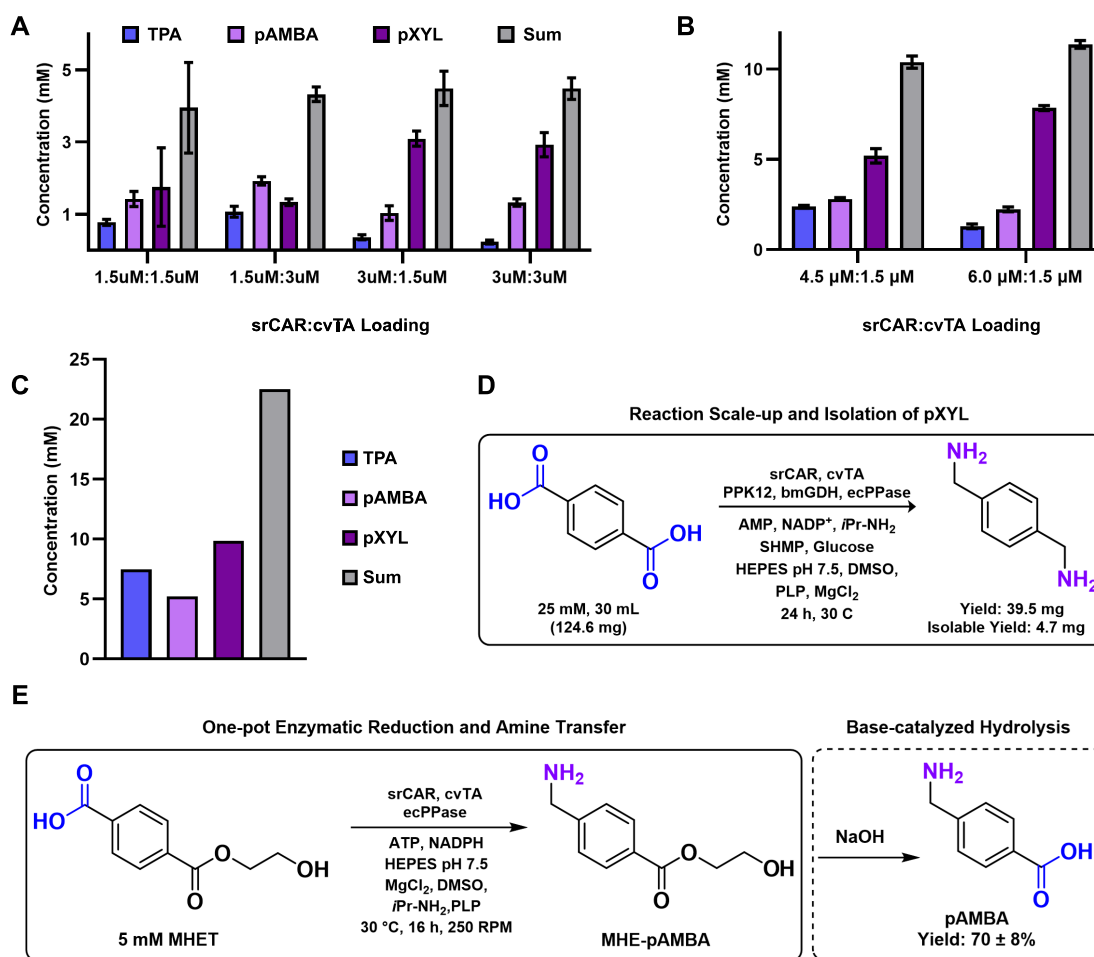


Figure 5. One-pot bioconversion of PET-derived carboxylates to amines. (A) One-pot in vitro bioconversion of TPA to pXYL by a five-enzyme cascade consisting of srCAR, cvTA, and a three-enzyme cofactor regeneration cascade yielded high reaction selectivity for amines and an appreciable yield of 62% of the desired diamine pXYL at 3 μ M srCAR loading. By tuning the ratio of CAR to TA, we saw a corresponding increase in yield and selectivity toward the diamine when the CAR concentration was increased. (B) At 10 mM loading of TPA as the substrate, we demonstrated 69 \pm 1% yield of TPA to pXYL in one pot at a srCAR/cvTA ratio of 6.0:1.5 μ M. (C) Operating at the same 6.0:1.5 μ M srCAR/cvTA ratio used in the 10 mM experiment, we scaled up the reaction to 30 mL volume at 25 mM substrate loading, which corresponds to 100 mg of theoretical yield of pXYL. We added 6.0 μ M srCAR, 1.5 μ M cvTA, 1.5 μ M bmGDH (tetramer basis), 0.84 mg/mL PPK12, and 1.5 μ M ecPPase (hexamer basis) to the pH-adjusted buffer that contained 100 mM HEPES pH 7.5, 1 mM AMP, 1 mM NADP⁺, 100 mM iPr-NH₂, 15 mM SHMP, 50 mM glucose, 5% v/v DMSO, 2 mM PLP, 30 mM MgCl₂, and 25 mM TPA. The reaction proceeded for 24 h at 30 °C and was mixed by inversion with a rotating rotisserie. After 24 h, endpoint HPLC showed 9.8 mM or 39.5 mg pXYL in solution. As expected, pAMBA was also present at 5.2 mM, and unreacted TPA comprised 7.5 mM of the reaction. This experiment was conducted in $N = 1$. (D) After performing a two-phase organic extraction, we isolated 4.7 mg of pXYL, even though the overall reaction yielded 39.5 mg, as quantified by HPLC. (E) Broad substrate specificity of srCAR and cvTA allowed us to produce the pharmaceutically relevant monoamine, pAMBA, solely by switching the starting substrate from TPA to MHET and performing a simple hydrolysis step to liberate pAMBA, yielding 70 \pm 8% of the desired product. For the bioconversion of MHET to pAMBA, 6.0 μ M srCAR, 1.5 μ M cvTA (dimer basis), 0.75 μ M ecPPase (hexamer basis), 6 mM NADPH, 6 mM ATP, 20 mM MgCl₂, 100 mM HEPES pH 7.5, 5% (v/v) DMSO, 20 mM iPr-NH₂, 400 μ M PLP, and 5 mM MHET were added to a total volume of 100 μ L. Following a 16 h incubation at 30 °C, the reaction was basified by NaOH to a final concentration of 0.5 M and analyzed using HPLC-UV. Sample size for panels (A,B,E) is $N = 3$. Error bars are shown as the mean \pm standard deviation.

correlation of our experimental results with the obtained $d1$ values with our docked AMP-acyl TPA. Interestingly, we observed binding of MHET in the predicted srCAR structure, indicating that the bulky ethylene glycol sidechain could be accommodated near position A315 in the binding pocket (Figure S7C). Overall, our computational analysis indicates that much remains to be understood about CAR specificity and that, for the time being, empirical studies such as ours that sample a wide range of CARs and potential substrates are essential toward elucidating the specificity of this important enzyme family.

Characterization of CAR Activity on PET Deconstruction Products with a Cofactor Regeneration Cascade.

Given the activity of CARs on several substrates of interest, we sought to provide higher substrate loading and monitor reaction progress over time. CARs are dependent on ATP and NADPH to catalyze the reduction of acids; however, supplementation of exogenous cofactors becomes more cost-prohibitive at higher substrate loading. For enzymatic ATP regeneration, we expressed a recently discovered type 2-III polyphosphate kinase (PPK2-III) from an unclassified *Erysipelotrichaceae* bacterium (PPK12) that was coupled to

CAR to produced aldehydes.⁴⁴ PPK12 converts AMP to ATP using sodium hexametaphosphate (SHMP) as a polyphosphate (polyP) source. Though initially we noticed our reaction stalling using this enzyme, we later discovered that polyP was inhibitory to either our CAR or PPK12 and that optimizing the polyP concentration allowed the CAR reaction to proceed (Figure S8). For NADPH regeneration, we expressed and tested the activity of an engineered glucose dehydrogenase from *Bacillus megaterium* (bmGDH) (Figure S9).⁶⁷ Lastly, the addition of purified inorganic pyrophosphatase from *E. coli* (ecPPase) eliminated accumulation of inorganic pyrophosphate (PP_i), which is inhibitory to CAR.⁶⁸

After verifying the activity of the key regeneration enzymes, we sequentially characterized the effect of bmGDH and PPK12 with srCAR. At our chosen endpoint of 24 h, we observed that the srCAR-catalyzed reduction of 5 mM TPA does not reach completion to TPAL, even with exogenous supplementation of both NADPH and ATP in excess as measured by HPLC-UV (Figures 4A, S10, and S11). However, we noticed that addition of bmGDH resulted in the formation of an overreduced product, 4-(hydroxymethyl)benzaldehyde (HMB), at 1 mM concentration of NADP⁺. The formation of alcohols is not known to be catalyzed by CAR⁴³ but has been reported in systems in which protein preparations may contain impurities.⁶⁹ Recently, the ability of some CARs to also catalyze overreduction of aldehydes to alcohols was shown.⁷⁰

To better understand the timescale of CAR-catalyzed reduction of PET-derived substrates, we performed a time-course analysis of srCAR-catalyzed reduction of 5 mM TPA, 4FBA, mmTPA, and MHET (Figure 4B). After 24 h, the conversion of TPA is 77%; however, the yield of the desired dialdehyde was only 39%, with 22% of the aldehyde products overreduced to HMB. Interestingly, by using 4FBA as the starting substrate, we see complete conversion of 5 mM 4FBA to 5 mM TPAL. The improved conversion was consistent with our heatmap analysis and our biochemical characterization of srCAR, which suggested that the lower substrate K_m of 4FBA increases the catalytic efficiency of the reaction. At later timepoints, we again saw the undesired production of HMB. Reduction of mmTPA to methyl 4-formylbenzoate (mm4FBA) by srCAR rapidly reached 65% conversion in 30 min but was likely subject to overreduction, as reported previously.⁷¹ Fortunately, we observed a potential alternate route to producing aldehydes from PET deconstruction products that mitigated overreduction. By supplementing 5 mM MHET to the multienzyme reduction cascade, we saw complete conversion of MHET within 4 h, with a yield of 85% of the desired product, 2-hydroxyethyl 4-formylbenzoate (MHE-4FBA).

To verify the viability of this potential alternative reductive pathway to produce aldehydes of interest, we sought to hydrolyze the ester group on both mm4FBA and MHE-4FBA to form 4FBA. srCAR can further reduce 4FBA to TPAL (Figure 4B). Lipases are well-suited for this application as they perform the hydrolysis of ester bonds in aqueous environments.⁷² Previously, lipase B from *Candida antarctica* (CALB) was used to deconstruct MHET to TPA.⁷³ Driven by the structural similarity of these carboxylates to their corresponding aldehydes, we used 1 U of CALB to perform the hydrolysis of 1 mM of both mm4FBA and MHE-4FBA, which yielded 4FBA as a sole product as quantified by HPLC-UV after 24 h incubation at 30 °C (Figure 4C). The ease of accessing diverse monoester derivatives of terephthalate from PET deconstruc-

tion coupled with the broad reduction activity of srCAR allows for the design of modular enzyme cascades to synthesize two distinct aldehydes, 4FBA or TPAL, and provides opportunities for downstream functionalization by cvTA.

One-Pot Synthesis of Diamine by a Reductive Biocatalytic Cascade. After determining the optimal CAR and ω -TA for use in our cascade, we developed a one-pot assay to produce our diamine of interest, pXYL. We combined srCAR, cvTA, and the 3-enzyme cofactor regeneration system in a one-pot assay with 5 mM TPA (Figure 5A, Supporting Information Methods). Using HPLC-UV (Figure S12, Supporting Information Methods), we obtained distinct retention times for all different potential reaction intermediates of our system. At 1.5 μ M loading of both srCAR and cvTA, we obtained a yield of 35% pXYL and 28% pAMBA, with no aldehyde species present during the reaction endpoint. Encouraged by this result, we looked at tuning the enzyme loading to see if we could shift the product distribution toward the diamine. Increasing the cvTA concentration from 1.5 to 3 μ M in the presence of 1.5 μ M srCAR slightly shifted the product yield to 38% pAMBA and 26% pXYL. Upon addition of 3 μ M srCAR, we saw a shift in distribution to favor the diamine. Our highest yield in this experiment was 62% yield of pXYL at a 3 μ M loading of srCAR and 1.5 μ M loading of cvTA. Coupling srCAR to cvTA resulted in no detectable formation of alcohols, which we observed when srCAR was coupled only to the cofactor regeneration cascade.

To test the cascade performance at higher substrate loading, we increased the loading of TPA to 10 mM. From our previous experiments, we saw that the increase in srCAR concentration drives the product distribution toward pXYL. We therefore further increased the concentration of srCAR to 4.5 and 6.0 μ M, keeping the concentration of cvTA at 1.5 μ M. We achieved a $69 \pm 1\%$ yield of our target diamine at 6.0 μ M srCAR and 1.5 μ M cvTA loading (Figure 5B). This result is unusually promising given the presence of only one observed byproduct, pAMBA, at 22% of the final reaction mix, the relatively low concentration of the enzyme, and the ability to use single variants of CAR and TA enzymes. Our enzyme cascade achieves a higher yield of desired diamine (69% instead of 30%) than a previously reported CAR/TA cascade that was supplied with 10 mM adipic acid.⁴⁰ There are several differences between these two efforts that are worth noting. Our system features aromatic compounds as opposed to medium-chain aliphatic compounds, thus generating aldehyde intermediates that differ with respect to reactivity and volatility. Our system for diamine production features the provision of lower-cost precursors to the required CAR cofactors (AMP and NADP⁺) instead of the actual CAR cofactors (ATP and NADPH). Our system also includes a different amine donor (we use *i*Pr-NH₂ instead of alanine) and no regeneration of the amine donor. Finally, our cascade features reduced complexity by identifying single CAR and TA variants capable of performing the bifunctional reduction and amination reactions. However, despite these differences in conditions, our enzyme concentrations were similar (6 μ M srCAR in our cascade vs 3 μ M each of two CAR variants).

Reaction Scaleup, Diamine Isolation, and Characterization. After successfully achieving a yield of 69% of the target diamine in 100 μ L reactions starting from 10 mM TPA, we scaled up our cascade to a 30 mL reaction volume at 25 mM TPA loading (124.6 mg of TPA, which if fully converted would result in 100 mg of pXYL). As before, we combined 6.0

μM srCAR with 1.5 μM cvTA along with the three-enzyme cofactor regeneration cascade but this time in a buffer that contained 100 mM *iPr*-NH₂ to maintain 4× molar excess of amine donor. The high concentration of the amine donor required us to acidify the buffer pH back down to 7.5. At the 24 h endpoint, we sampled a small volume (100 μL) for analysis by HPLC. Encouragingly, we found that we obtained 9.8 mM pXYL (39.5 mg pXYL, or 40% yield) along with 5.2 mM pAMBA and 7.5 mM remaining TPA (Figure 5C). To selectively isolate pXYL, we quenched the remaining volume with 10 M HCl, passed the reaction mix through a Buchner funnel, and basified the filtrate with 10 N NaOH. We then performed two 15 mL extractions with dichloromethane (DCM) as the organic phase. Both DCM extractions resulted in the formation of emulsions, to which we added a saturated brine solution. Due to the difficulties encountered during the extraction, we achieved an isolable yield of only 4.7 mg (Figure 5D). Using HPLC, we observed the presence of pXYL remaining in the aqueous phase after extraction (Figure S13A). However, through ¹H NMR, we were able to verify the identity of the extracted diamine (Figure S13B). Overall, the reaction yield that we obtained of 40% is promising for the possibility of scaleup; however, additional investigation of recovery techniques will be needed to utilize PET-derived pXYL in polymer applications.

One-Pot Synthesis of a Monoamine by a Reductive Biocatalytic Cascade. Lastly, we took advantage of the wide substrate scope of both srCAR and cvTA to design an enzyme cascade for the synthesis of monofunctionalized amine, pAMBA, an antifibrinolytic drug.²⁶ We had previously observed selective reduction of MHET to MHE-4FBA by srCAR (Figures 3D and 4B). Owing to the wide substrate scope of cvTA and the previous result that cvTA is highly active on a structurally similar aldehyde, 4FBA, we posited that MHE-4FBA can also be accepted as a substrate by cvTA. To experimentally confirm cvTA activity on MHE-4FBA, we added 5 mM of chemically synthesized substrate to 1.5 μM cvTA and 20 mM *iPr*-NH₂. Because our substrate was not pure and because we did not have access to the monoamine product anticipated prior to ester hydrolysis, we used a different approach to monitor the reaction progress. We observed depletion in the substrate peak and formation of a product peak on HPLC at 24 h (Figure S14A). To verify that the product peak corresponded to an amine, we performed OPA derivatization, which specifically reacts with amines, and observed a new peak on the chromatogram (Figure S14B). Encouraged by this result, we then performed a one-pot conversion of 5 mM MHET by adding 6.0 μM srCAR and 1.5 μM cvTA, followed by base-catalyzed hydrolysis of the glycol ester. This reaction produced pAMBA at 70 ± 8% yield, with high selectivity and no diamine formation (Figure 5E). The wide substrate scope of CAR and ω -TA allows for a modular route to upcycled two distinct amine products with high selectivity, depending on the available PET-derived starting substrate.

CONCLUSIONS

New technologies must be developed to combat the growing amount of plastic waste, and chemical deconstruction methods alone may not sufficiently incentivize nor diversify the recycling of plastic waste. Biocatalytic synthesis of diamines and monoamines from PET-derived esters or carboxylic acid monomers offers an opportunity to upcycle PET for

applications in materials and therapeutics. Given the selectivity of enzymes, one would expect that the enzymes in our cascade would be amenable to relatively seamless integration with enzymatic deconstruction approaches, such as those that rely on cutinases or PETases. In principle, this integration could also occur in one pot, increasing the attractiveness and cost-competitiveness of biocatalytic valorization strategies. Additionally, because the aromatic character of the starting material is preserved and only a small number of molecular transformations is required, the routes to these products offer higher theoretical yields than when starting from traditional fermentative carbon sources such as glucose.

This study revealed several interesting aspects of substrate specificity for three industrially relevant enzyme classes. We first established that cvTA could catalyze amine transfer to the PET-derived aldehydes 4FBA and TPAL, which resulted in a high yield of the desired amines pXYL and pAMBA, respectively. With this knowledge, we then explored enzymatic synthesis of aldehydes from PET-derived substrates. Through a bioprospecting approach, we identified that srCAR could catalyze the bifunctional enzymatic reduction of TPA to TPAL (via 4FBA), which is a relatively newly observed phenomenon. Moreover, it was encouraging that this single CAR could accept MHET and mmTPA as substrates for reduction to their respective monoaldehyde products, which we show could be de-esterified enzymatically by a lipase. Our bioprospecting approach also revealed potential utility of a previously uncharacterized putative aryl acid reductase from *T. reesei*, whose activity and substrate scope were favorable and merit further testing.

By pairing enzymatic aldehyde production to amine transfer, we developed a one-pot route to produce both diamines and monoamines. Upon addition of 10 mM TPA to srCAR, cvTA, and enzymes for cofactor regeneration, we achieved 69 ± 1% of the desired diamine pXYL. Encouraged by the efficiency of the cascade, we scaled up our reaction to 30 mL and produced pXYL at 40% yield. Reaction scaleup and optimization continue to be an area of interest as we believe our results pave the way for demonstrations of the utility of pXYL in polymeric materials applications. We also highlight the ability of our chosen enzymes to accept multiple starting substrates depending on the desired product output. This modularity is highlighted in the one-pot enzymatic reduction and amine transfer of MHET, which, followed by base-catalyzed ester hydrolysis, produced the monoamine therapeutic pAMBA at high selectivity and yield of 70 ± 8% from 5 mM of the starting substrate. Given that only micromolar enzyme concentrations were used, we believe this cascade has the potential for polymer and medicinal syntheses, where larger quantities and purities of amine products would be required. Overall, our reductive cascade design aids in further incentivizing the diversion of PET waste from the environment by augmenting the sparse list of biological PET upcycling strategies.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acscatal.2c06219>.

Supplementary figures, supplementary methods, cloning and purification, HPLC methods, chemical synthesis, and supplementary tables (PDF)

AUTHOR INFORMATION

Corresponding Author

Aditya M. Kunjapur – Department of Chemical & Biomolecular Engineering, University of Delaware, Newark, Delaware 19716, United States; Center for Plastics Innovation, University of Delaware, Newark, Delaware 19716, United States; orcid.org/0000-0001-6869-9530; Email: kunjapur@udel.edu

Authors

Madan R. Gopal – Department of Chemical & Biomolecular Engineering, University of Delaware, Newark, Delaware 19716, United States; Center for Plastics Innovation, University of Delaware, Newark, Delaware 19716, United States

Roman M. Dickey – Department of Chemical & Biomolecular Engineering, University of Delaware, Newark, Delaware 19716, United States; Center for Plastics Innovation, University of Delaware, Newark, Delaware 19716, United States

Neil D. Butler – Department of Chemical & Biomolecular Engineering, University of Delaware, Newark, Delaware 19716, United States

Michael R. Talley – Department of Chemistry and Biochemistry and Center for Plastics Innovation, University of Delaware, Newark, Delaware 19716, United States

Daniel T. Nakamura – Center for Plastics Innovation, University of Delaware, Newark, Delaware 19716, United States

Ashlesha Mohapatra – Department of Chemical & Biomolecular Engineering, University of Delaware, Newark, Delaware 19716, United States

Mary P. Watson – Department of Chemistry and Biochemistry and Center for Plastics Innovation, University of Delaware, Newark, Delaware 19716, United States; orcid.org/0000-0002-1879-5257

Wilfred Chen – Department of Chemical & Biomolecular Engineering, University of Delaware, Newark, Delaware 19716, United States; Center for Plastics Innovation, University of Delaware, Newark, Delaware 19716, United States; orcid.org/0000-0002-6386-6958

Complete contact information is available at: <https://pubs.acs.org/10.1021/acscatal.2c06219>

Author Contributions

M.R.G. performed cloning and purification, designed all CAR and ω -TA experiments, one-pot experiments, and scaleup, and wrote the manuscript. R.M.D. performed cloning and purification, conducted lipase experiments and molecular docking of CAR A-domains, and contributed to writing the manuscript. N.D.B. designed the SSN and contributed to writing the manuscript. M.R.T. synthesized standards and contributed to writing the Supporting Information. D.T.N. isolated pXYL and assisted M.R.G. with scaleup. A.M. assisted M.R.G. in testing one-pot enzymatic bioconversion conditions. A.M.K. conceptualized the overall goals of this study and contributed to writing the manuscript. A.M.K., W.C., and M.P.W. secured funding, provided oversight, and reviewed the manuscript.

Funding

A.M.K., W.C., M.P.W., M.R.G., R.M.D., and M.R.T. acknowledge the Center for Plastics Innovation, an Energy Frontier

Research Center funded by the U.S. Department of Energy, Office of Science, Basic Energy Sciences (grant number DESC0021166). Cloning work was performed under proposal 506446 by the U.S. Department of Energy Joint Genome Institute, a DOE Office of Science User Facility, supported by the Office of Science of the U.S. Department of Energy operated under contract no. DE-AC02-05CH11231. More information can be found at [10.46936/10.25585/60001335](https://doi.org/10.46936/10.25585/60001335) and <https://ror.org/04xm1d337>.

Notes

The authors declare the following competing financial interest(s): A.M.K., M.R.G., R.M.D., and W.C. have filed for a provisional patent related to this work.

ACKNOWLEDGMENTS

The authors would like to thank the members of the University of Delaware Center for Plastics Innovation for guidance and support on this project.

ABBREVIATIONS

CAR, carboxylic acid reductase; ω -TA, ω -transaminase; PET, polyethylene terephthalate; TPA, terephthalic acid; 4FBA, 4-formylbenzoic acid; TPAL, terephthalaldehyde; pAMBA, *para*-(aminomethyl)benzoic acid; pAMB, *para*-(aminomethyl)-benzaldehyde; pXYL, *para*-xylylenediamine; HMB, 4-(hydroxymethyl)benzaldehyde; MHE-4FBA, 2-hydroxyethyl 4-formylbenzoate; mm4FBA, methyl 4-formylbenzoate; BHET, bis(2-hydroxyethyl) terephthalate; MHET, mono(2-hydroxyethyl) terephthalic acid; NADPH, nicotinamide adenine dinucleotide phosphate (reduced); NADP⁺, nicotinamide adenine dinucleotide phosphate (oxidized); ATP, adenosine 5'-triphosphate; AMP, adenosine 5'-monophosphate; SHMP, sodium hexametaphosphate

REFERENCES

- (1) Altman, S.; Rosetto, G. Achieving a Circular Bioeconomy for Plastics. *Science* **2021**, *373*, 47–49.
- (2) Mitrano, D. M.; Wagner, M. A Sustainable Future for Plastics Considering Material Safety and Preserved Value. *Nat. Rev. Mater.* **2022**, *7*, 71–73.
- (3) Korley, L. T. J.; Epps, T. H.; Helms, B. A.; Ryan, A. J. Toward Polymer Upcycling-Adding Value and Tackling Circularity. *Science* **2021**, *373*, 66–69.
- (4) Fagnani, D. E.; Tami, J. L.; Copley, G.; Clemons, M. N.; Getzler, Y. D. Y. L.; McNeil, A. J. 100th Anniversary of Macromolecular Science Viewpoint: Redefining Sustainable Polymers. *ACS Macro Lett.* **2021**, *10*, 41–53.
- (5) Zhao, X.; Boruah, B.; Chin, K. F.; Đokić, M.; Modak, J. M.; Soo, H. Upcycling to Sustainably Reuse Plastics. *Adv. Mater.* **2022**, *34*, 2100843.
- (6) Rahimi, A. R.; García, J. M. Chemical Recycling of Waste Plastics for New Materials Production. *Nat. Rev. Chem.* **2017**, *1*, 1–11.
- (7) Sardon, H.; Dove, A. P. Plastics Recycling with a Difference. *Science* **2018**, *360*, 380–381.
- (8) Christensen, P. R.; Scheuermann, A. M.; Loeffler, K. E.; Helms, B. A. Closed-Loop Recycling of Plastics Enabled by Dynamic Covalent Diketoneamine Bonds. *Nat. Chem.* **2019**, *11*, 442–448.
- (9) Payne, J.; Jones, M. D. The Chemical Recycling of Polyesters for a Circular Plastics Economy: Challenges and Emerging Opportunities. *ChemSusChem* **2021**, *14*, 4041–4070.
- (10) Yue, Q. F.; Wang, C. X.; Zhang, L. N.; Ni, Y.; Jin, Y. X. Glycolysis of Poly(Ethylene Terephthalate) (PET) Using Basic Ionic Liquids as Catalysts. *Polym. Degrad. Stab.* **2011**, *96*, 399–403.

- (11) Wang, L.; Nelson, G. A.; Toland, J.; Holbrey, J. D. Glycolysis of PET Using 1,3-Dimethylimidazolium-2-Carboxylate as an Organocatalyst. *ACS Sustainable Chem. Eng.* **2020**, *8*, 13362–13368.
- (12) Wang, H.; Liu, Y.; Li, Z.; Zhang, X.; Zhang, S.; Zhang, Y. Glycolysis of Poly(Ethylene Terephthalate) Catalyzed by Ionic Liquids. *Eur. Polym. J.* **2009**, *45*, 1535–1544.
- (13) Karayannidis, G. P.; Chatziavagoustis, A. P.; Achilias, D. S. Poly(Ethylene Terephthalate) Recycling and Recovery of Pure Terephthalic Acid by Alkaline Hydrolysis. *Adv. Polym. Technol.* **2002**, *21*, 250–259.
- (14) Brown, G. E.; O'Brien, R. C. Method For Recovering Terephthalic Acid And Ethylene Glycol From Polyester Materials. U.S. Patent 3,952,053 A, April 20, 1974.
- (15) Müller, R. J.; Schrader, H.; Profe, J.; Dresler, K.; Deckwer, W. D. Enzymatic Degradation of Poly(Ethylene Terephthalate): Rapid Hydrolyse Using a Hydrolase from *T. Fusca*. *Macromol Rapid Commun* **2005**, *26*, 1400–1405.
- (16) Yoshida, S.; Hiraga, K.; Takehana, T.; Taniguchi, I.; Yamaji, H.; Maeda, Y.; Toyohara, K.; Miyamoto, K.; Kimura, Y.; Oda, K. A Bacterium That Degrades and Assimilates Poly(Ethylene Terephthalate). *Science* **2016**, *351*, 1196–1199.
- (17) Lu, H.; Diaz, D. J.; Czarnecki, N. J.; Zhu, C.; Kim, W.; Shroff, R.; Acosta, D. J.; Alexander, B. R.; Cole, H. O.; Zhang, Y.; Lynd, N. A.; Ellington, A. D.; Alper, H. S. Machine Learning-Aided Engineering of Hydrolases for PET Depolymerization. *Nature* **2022**, *604*, 662–667.
- (18) Kim, H. T.; Kim, J. K.; Cha, H. G.; Kang, M. J.; Lee, H. S.; Khang, T. U.; Yun, E. J.; Lee, D. H.; Song, B. K.; Park, S. J.; Joo, J. C.; Kim, K. H. Biological Valorization of Poly(Ethylene Terephthalate) Monomers for Upcycling Waste PET. *ACS Sustainable Chem. Eng.* **2019**, *7*, 19396–19406.
- (19) Werner, A. Z.; Clare, R.; Mand, T. D.; Pardo, I.; Ramirez, K. J.; Haugen, S. J.; Bratti, F.; Dexter, G. N.; Elmore, J. R.; Huenemann, J. D.; Peabody, G. L.; Johnson, C. W.; Rorrer, N. A.; Salvachúa, D.; Guss, A. M.; Beckham, G. T. Tandem Chemical Deconstruction and Biological Upcycling of Poly(Ethylene Terephthalate) to β -Ketoadipic Acid by *Pseudomonas Putida* KT2440. *Metab. Eng.* **2021**, *67*, 250–261.
- (20) Sullivan, K. P.; Werner, A. Z.; Ramirez, K. J.; Ellis, L. D.; Bussard, J. R.; Black, B. A.; Brandner, D. G.; Bratti, F.; Buss, B. L.; Dong, X.; Haugen, S. J.; Ingraham, M. A.; Konev, M. O.; Michener, W. E.; Miscall, J.; Pardo, I.; Woodworth, S. P.; Guss, A. M.; Román-Leshkov, Y.; Stahl, S. S.; Beckham, G. T. Mixed Plastics Waste Valorization through Tandem Chemical Oxidation and Biological Funneling. *Science* **2022**, *378*, 207–211.
- (21) Dickey, R. M.; Forti, A. M.; Kunjapur, A. M. Advances in Engineering Microbial Biosynthesis of Aromatic Compounds and Related Compounds. *Bioresour. Bioprocess* **2021**, *8*, 1–17.
- (22) Nanclares, J.; Petrović, Z. S.; Javni, L.; Ionescu, M.; Jaramillo, F. Segmented Polyurethane Elastomers by Nonisocyanate Route. *J. Appl. Polym. Sci.* **2015**, *132*, 42492.
- (23) Stavila, E.; Alberda van Ekenstein, G. O. R.; Loos, K. Enzyme-Catalyzed Synthesis of Aliphatic-Aromatic Oligoamides. *Biomacromolecules* **2013**, *14*, 1600–1606.
- (24) Process for the Manufacture of Nitriles and Their Corresponding Amines. CN 108164381 B, March 16, 2018.
- (25) Blackmon, K. P.; Fox, D. W.; Shafer, S. J. Process for Converting Pet Scrap to Diamine Monomers. U.S. Patent 4,973,746 A, November 27, 1990.
- (26) Banach, L.; Janczewski, L.; Kajdanek, J.; Milowska, K.; Kolodziejczyk-Czepas, J.; Galita, G.; Rozpedek-Kaminska, W.; Kucharska, E.; Majsterek, L.; Kolesinska, B. Synthesis and Hemostatic Activity of New Amide Derivatives. *Molecules* **2022**, *27*, 2271.
- (27) Dörwald, F. Zaragoza. Monofunctionalization of Symmetric Difunctional Substrates. In *Side Reactions in Organic Synthesis: A Guide to Successful Synthesis Design*; Wiley-VCH, 2005; pp 333–353.
- (28) Bell, E. L.; Finnigan, W.; France, S. P.; Green, A. P.; Hayes, M. A.; Hepworth, L. J.; Lovelock, S. L.; Niikura, H.; Osuna, S.; Romero, E.; Ryan, K. S.; Turner, N. J.; Flitsch, S. L. Biocatalysis. *Nat. Rev. Methods Primers* **2021**, *1*, 46.
- (29) Prather, K. L. J.; Martin, C. H. De Novo Biosynthetic Pathways: Rational Design of Microbial Chemical Factories. *Curr. Opin. Biotechnol.* **2008**, *19*, 468–474.
- (30) Turner, N. J.; O'Reilly, E. Biocatalytic Retrosynthesis. *Nat. Chem. Biol.* **2013**, *9*, 285–288.
- (31) Steffen-Munsberg, F.; Vickers, C.; Thontowi, A.; Schätzle, S.; Meinhardt, T.; Svedendahl Humble, M.; Land, H.; Berglund, P.; Bornscheuer, U. T.; Höhne, M. Revealing the Structural Basis of Promiscuous Amine Transaminase Activity. *ChemCatChem* **2013**, *5*, 154–157.
- (32) Shin, J.-S.; Kim, B.-G. Kinetic Modeling Of-Transamination for Enzymatic Kinetic Resolution of-Methylbenzylamine. *Biotechnol. Bioeng.* **1998**, *60*, 534–540.
- (33) Kaulmann, U.; Smithies, K.; Smith, M. E. B.; Hailes, H. C.; Ward, J. M. Substrate Spectrum of ω -Transaminase from *Chromobacterium Violaceum* DSM30191 and Its Potential for Biocatalysis. *Enzyme Microb. Technol.* **2007**, *41*, 628–637.
- (34) Khusnutdinova, A. N.; Flick, R.; Popovic, A.; Brown, G.; Tchigvintsev, A.; Nocek, B.; Correia, K.; Joo, J. C.; Mahadevan, R.; Yakunin, A. F. Exploring Bacterial Carboxylate Reductases for the Reduction of Bifunctional Carboxylic Acids. *Biotechnol. J.* **2017**, *12*, 1600751.
- (35) Winkler, M. Carboxylic Acid Reductase Enzymes (CARs). *Curr. Opin. Chem. Biol.* **2018**, *43*, 23–29.
- (36) Bayer, T.; Pfaff, L.; Branson, Y.; Becker, A.; Wu, S.; Bornscheuer, U. T.; Wei, R. Biosensor and Chemo-Enzymatic One-Pot Cascade Applications to Detect and Transform PET-Derived Terephthalic Acid in Living Cells. *iScience* **2022**, *25*, 104326.
- (37) Butler, N.; Kunjapur, A. M. Carboxylic Acid Reductases in Metabolic Engineering. *J. Biotechnol.* **2020**, *307*, 1–14.
- (38) Yuan, S.; Jin, M.; Xu, C.; Yin, W.; Zhao, M.; Xian, M.; Liu, W. Self-Sufficient Whole-Cell Biocatalysis for 3-(Aminomethyl) Pyridine Synthesis. *Biochem. Eng. J.* **2022**, *183*, 108457.
- (39) France, S. P.; Hussain, S.; Hill, A. M.; Hepworth, L. J.; Howard, R. M.; Mulholland, K. R.; Flitsch, S. L.; Turner, N. J. One-Pot Cascade Synthesis of Mono- and Disubstituted Piperidines and Pyrrolidines Using Carboxylic Acid Reductase (CAR), ω -Transaminase (ω -TA), and Imine Reductase (IREd) Biocatalysts. *ACS Catal.* **2016**, *6*, 3753–3759.
- (40) Fedorchuk, T. P.; Khusnutdinova, A. N.; Evdokimova, E.; Flick, R.; Di Leo, R.; Stogios, P.; Savchenko, A.; Yakunin, A. F. One-Pot Biocatalytic Transformation of Adipic Acid to 6-Aminocaproic Acid and 1,6-Hexamethylenediamine Using Carboxylic Acid Reductases and Transaminases. *J. Am. Chem. Soc.* **2020**, *142*, 1038–1048.
- (41) Chen, S.; Land, H.; Berglund, P.; Humble, M. S. Stabilization of an Amine Transaminase for Biocatalysis. *J. Mol. Catal. B: Enzym.* **2016**, *124*, 20–28.
- (42) Savile, C. K.; Janey, J. M.; Mundorff, E. C.; Moore, J. C.; Tam, S.; Jarvis, W. R.; Colbeck, J. C.; Krebber, A.; Fleitz, F. J.; Brands, J.; Devine, P. N.; Huisman, G. W.; Hughes, G. J. Biocatalytic Asymmetric Synthesis of Chiral Amines from Ketones Applied to Sitagliptin Manufacture. *Science* **2010**, *329*, 305–309.
- (43) Gahlthorn, D.; Dunstan, M. S.; Quaglia, D.; Klumbys, E.; Lockhart-Cairns, M. P.; Hill, A. M.; Derrington, S. R.; Scrutton, N. S.; Turner, N. J.; Leys, D. Structures of Carboxylic Acid Reductase Reveal Domain Dynamics Underlying Catalysis. *Nat. Chem. Biol.* **2017**, *13*, 975–981.
- (44) Tavanti, M.; Hosford, J.; Lloyd, R. C.; Brown, M. J. B. ATP Regeneration by a Single Polyphosphate Kinase Powers Multigram-Scale Aldehyde Synthesis in Vitro. *Green Chem.* **2021**, *23*, 828–837.
- (45) Gerlt, J. A.; Bouvier, J. T.; Davidson, D. B.; Imker, H. J.; Sadkhin, B.; Slater, D. R.; Whalen, K. L. Enzyme Function Initiative-Enzyme Similarity Tool (EFI-EST): A Web Tool for Generating Protein Sequence Similarity Networks. *Biochim. Biophys. Acta, Proteomics* **2015**, *1854*, 1019–1037.
- (46) Stolterfoht, H.; Schwendenwein, D.; Sensen, C. W.; Rudroff, F.; Winkler, M. Four Distinct Types of E.C. 1.2.1.30 Enzymes Can

- Catalyze the Reduction of Carboxylic Acids to Aldehydes. *J. Biotechnol.* **2017**, *257*, 222–232.
- (47) He, A.; Li, T.; Daniels, L.; Fotheringham, I.; Rosazza, J. P. N. Nocardia Sp. Carboxylic Acid Reductase: Cloning, Expression, and Characterization of a New Aldehyde Oxidoreductase Family. *Appl. Environ. Microbiol.* **2004**, *70*, 1874–1881.
- (48) Li, C.; Matsuda, Y.; Gao, H.; Hu, D.; Yao, X. S.; Abe, I. Biosynthesis of LL-Z1272 β : Discovery of a New Member of NRPS-like Enzymes for Aryl-Aldehyde Formation. *ChemBioChem* **2016**, *17*, 904–907.
- (49) Schwendenwein, D.; Fiume, G.; Weber, H.; Rudroff, F.; Winkler, M. Selective Enzymatic Transformation to Aldehydes in Vivo by Fungal Carboxylate Reductase from Neurospora Crassa. *Adv. Synth. Catal.* **2016**, *358*, 3414–3421.
- (50) Horvat, M.; Fiume, G.; Fritsche, S.; Winkler, M. Discovery of Carboxylic Acid Reductase (CAR) from Thermothelomyces Thermophila and Its Evaluation for Vanillin Synthesis. *J. Biotechnol.* **2019**, *304*, 44–51.
- (51) Horvat, M.; Fritsche, S.; Kourist, R.; Winkler, M. Characterization of Type IV Carboxylate Reductases (CARs) for Whole Cell-Mediated Preparation of 3-Hydroxytyrosol. *ChemCatChem* **2019**, *11*, 4171–4181.
- (52) Shannon, P.; Markiel, A.; Ozier, O.; Baliga, N. S.; Wang, J. T.; Ramage, D.; Amin, N.; Schwikowski, B.; Ideker, T. Cytoscape: A Software Environment for Integrated Models of Biomolecular Interaction Networks. *Genome Res.* **2003**, *13*, 2498–2504.
- (53) Sophos, N. A.; Vasiliou, V. Aldehyde Dehydrogenase Gene Superfamily: The 2002 Update. *Chem.-Biol. Interact.* **2003**, *143*–144, 5–22.
- (54) Guo, S.; Bhattacharjee, J. K. Posttranslational Activation, Site-Directed Mutation and Phylogenetic Analyses of the Lysine Biosynthesis Enzymes α -Aminoadipate Reductase Lyslp (AAR) and the Phosphopantetheinyl Transferase Lys7p (PPTase) from Schizosaccharomyces Pombe. *Yeast* **2004**, *21*, 1279–1288.
- (55) Venkitasubramanian, P.; Daniels, L.; Rosazza, J. P. N. Reduction of Carboxylic Acids by Nocardia Aldehyde Oxidoreductase Requires a Phosphopantetheinylated Enzyme. *J. Biol. Chem.* **2007**, *282*, 478–485.
- (56) Finnigan, W.; Thomas, A.; Cromar, H.; Gough, B.; Snajdrova, R.; Adams, J. P.; Littlechild, J. A.; Harmer, N. J. Characterization of Carboxylic Acid Reductases as Enzymes in the Toolbox for Synthetic Chemistry. *ChemCatChem* **2017**, *9*, 1005–1017.
- (57) Kalb, D.; Lackner, G.; Hoffmeister, D. Functional and Phylogenetic Divergence of Fungal Adenylate-Forming Reductases. *Appl. Environ. Microbiol.* **2014**, *80*, 6175–6183.
- (58) Kurokawa, H.; Ohshima, M. A.; Sugiyama, K.; Miura, H. Methanolysis of Polyethylene Terephthalate (PET) in the Presence of Aluminium Triisopropoxide Catalyst to Form Dimethyl Terephthalate and Ethylene Glycol. *Polym. Degrad. Stab.* **2003**, *79*, 529–533.
- (59) Ward, L. C.; McCue, H. v.; Rigden, D. J.; Kershaw, N. M.; Ashbrook, C.; Hatton, H.; Goulding, E.; Johnson, J. R.; Carnell, A. J. Carboxyl Methyltransferase Catalysed Formation of Mono- and Dimethyl Esters under Aqueous Conditions: Application in Cascade Biocatalysis. *Angew. Chem., Int. Ed.* **2022**, *61*, 1–8.
- (60) Knott, B. C.; Erickson, E.; Allen, M. D.; Gado, J. E.; Graham, R.; Kearns, F. L.; Pardo, I.; Topuzlu, E.; Anderson, J. J.; Austin, H. P.; Dominick, G.; Johnson, C. W.; Rorrer, N. A.; Szostkiewicz, C. J.; Copié, V.; Payne, C. M.; Woodcock, H.; Donohoe, B. S.; Beckham, G. T.; McGeehan, J. E. Characterization and Engineering of a Two-Enzyme System for Plastics Depolymerization. *Proc. Natl. Acad. Sci.* **2020**, *117*, 25476–25485.
- (61) Jumper, J.; Evans, R.; Pritzel, A.; Green, T.; Figurnov, M.; Ronneberger, O.; Tunyasuvunakool, K.; Bates, R.; Žídek, A.; Potapenko, A.; Bridgland, A.; Meyer, C.; Kohl, S. A. A.; Ballard, A. J.; Cowie, A.; Romera-Paredes, B.; Nikolov, S.; Jain, R.; Adler, J.; Back, T.; Petersen, S.; Reiman, D.; Clancy, E.; Zielinski, M.; Steinegger, M.; Pacholska, M.; Berghammer, T.; Bodenstein, S.; Silver, D.; Vinyals, O.; Senior, A. W.; Kavukcuoglu, K.; Kohli, P.; Hassabis, D. Highly Accurate Protein Structure Prediction with AlphaFold. *Nature* **2021**, *596*, 583–589.
- (62) Trott, O.; Olson, A. J. AutoDock Vina: Improving the Speed and Accuracy of Docking with a New Scoring Function, Efficient Optimization, and Multithreading. *J. Comput. Chem.* **2009**, *31*, 455–461.
- (63) Madeira, F.; Pearce, M.; Tivey, A. R. N.; Basutkar, P.; Lee, J.; Edbali, O.; Madhusoodanan, N.; Kolesnikov, A.; Lopez, R. Search and Sequence Analysis Tools Services from EMBL-EBI in 2022. *Nucleic Acids Res.* **2022**, *50*, W276–W279.
- (64) Jurcik, A.; Bednar, D.; Byska, J.; Marques, S. M.; Furmanova, K.; Daniel, L.; Kokkonen, P.; Brezovsky, J.; Strnad, O.; Stourac, J.; Pavelka, A.; Manak, M.; Damborsky, J.; Kozlikova, B. CAVER Analyst 2.0: Analysis and Visualization of Channels and Tunnels in Protein Structures and Molecular Dynamics Trajectories. *Bioinformatics* **2018**, *34*, 3586–3588.
- (65) Wang, L.; Sun, Y.; Diao, S.; Jiang, S.; Wang, H.; Wei, D. Rational Hinge Engineering of Carboxylic Acid Reductase from Mycobacterium Smegmatis Enhances Its Catalytic Efficiency in Biocatalysis. *Biotechnol. J.* **2022**, *17*, 2100441.
- (66) Qu, G.; Fu, M.; Zhao, L.; Liu, B.; Liu, P.; Fan, W.; Ma, J. A.; Sun, Z. Computational Insights into the Catalytic Mechanism of Bacterial Carboxylic Acid Reductase. *J. Chem. Inf. Model.* **2019**, *59*, 832–841.
- (67) Dirkmann, M.; Nowack, J.; Schulz, F. An in Vitro Biosynthesis of Sesquiterpenes Starting from Acetic Acid. *ChemBioChem* **2018**, *19*, 2146–2151.
- (68) Kunjapur, A. M.; Cervantes, B.; Prather, K. L. J. Coupling Carboxylic Acid Reductase to Inorganic Pyrophosphatase Enhances Cell-Free in Vitro Aldehyde Biosynthesis. *Biochem. Eng. J.* **2016**, *109*, 19–27.
- (69) Schwarz, A.; Hecko, S.; Rudroff, F.; Kohrt, J. T.; Howard, R. M.; Winkler, M. Cell-Free in Vitro Reduction of Carboxylates to Aldehydes: With Crude Enzyme Preparations to a Key Pharmaceutical Building Block. *Biotechnol. J.* **2021**, *16*, 2000315.
- (70) Daniel, B.; Hashem, C.; Leithold, M.; Sagmaister, T.; Tripp, A.; Stolterfoht-Stock, H.; Messenlehner, J.; Keegan, R.; Winkler, C. K.; Ling, J. G.; Younes, S. H. H.; Oberdorfer, G.; Abu Bakar, F. D.; Gruber, K.; Pavkov-Keller, T.; Winkler, M. Structure of the Reductase Domain of a Fungal Carboxylic Acid Reductase and Its Substrate Scope in Thioester and Aldehyde Reduction. *ACS Catal.* **2022**, *12*, 15668–15674.
- (71) Strohmeier, G. A.; Schwarz, A.; Andexer, J. N.; Winkler, M. Co-Factor Demand and Regeneration in the Enzymatic One-Step Reduction of Carboxylates to Aldehydes in Cell-Free Systems. *J. Biotechnol.* **2020**, *307*, 202–207.
- (72) Borrelli, G. M.; Trono, D. Recombinant Lipases and Their Use as Biocatalysts for Industrial Applications. *Int. J. Mol. Sci.* **2015**, *16*, 20774–20840.
- (73) Carniel, A.; Valoni, É.; Nicomedes, J.; Gomes, A. da C.; Castro, A. M. de. Lipase from Candida Antarctica (CALB) and Cutinase from Humicola Insolens Act Synergistically for PET Hydrolysis to Terephthalic Acid. *Process Biochem.* **2017**, *59*, 84–90.